


<b>P-065</b>	<b>Chitosan/alginate pellets with rutin developed for inflammatory bowel disease treatment</b>	
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## INTRODUCTION AND OBJECTIVES

Inflammatory bowel disease (IBD) is a refractory, chronic, recurrent and non-specific inflammatory disorder of intestinal tract. Rutin and other flavonoids exhibit anti-oxidant and anti-inflammatory effects; therefore they are the subject of interest as potential drugs for the treatment of several diseases including IBD (Rabiskova 2009). In our experiment, pellets based on chitosan/alginate for rutin colon delivery were prepared, evaluated *in vitro* and *in vivo* in experimental colitis in rats.

## MATERIALS AND METHODS

### Materials

Rutin (Sigma-Aldrich, Germany) as an active ingredient, chitosan (deacetylation degree of 91 %; JBiChem, China) and microcrystalline cellulose Avicel<sup>®</sup> PH 101 (FMC, Ireland) as fillers, and sodium alginate (Sigma-Aldrich, Germany) as the main coating excipient were used to prepare coated pellets. Acetate or phosphate buffers (pH 3.0, 6.8, 7.5, 4.0, 6.0, Ph. Eur. 6) and  $\beta$ -glucosidase (3811U/mg; MP Biomedicals, USA) were used for dissolution studies. Trinitrobenzensulfonic acid, hexadecyltrimethylammonium bromide (Fluka Biochemika, Germany) and o-dianisidine dihydrochloride (Sigma-Aldrich, Germany) were used in *in vivo* experiments.

### Pellet preparation

Pellets containing 30% of rutin, 45% of chitosan and 25% of microcrystalline cellulose were prepared using extrusion/ spheronization method (Pharmex 35T, Wyss & Probst, Germany), subsequently coated with alginate/chitosan (95:5) solution in Wurster type fluid bed unit (M-100, Medipo, Czech Republic) up to 18 % of total pellets mass, and dried in ventilated oven (Horo, Germany) at 40 °C for 3 hours.

### Pellet characterization

Pellet characteristics (mean diameter, sphericity, hardness, friability, Hausner ratio) were determined (Scala-Bertola 2009). Rutin content was measured spectrophotometrically at a wavelength of 360 nm (phosphate buffer pH 6.8; Lambda 25, Perkin Elmer Instruments, USA). Drug release was evaluated (1000 mL, 100 rpm, 37 °C, 360 nm; Sotax AT 7 Smart on-line, Donau Lab, Switzerland) using basket dissolution method with changing pH values simulating gastrointestinal tract conditions.  $\beta$ -glucosidase was added to the phosphate

buffer of pH 6.0 to decompose chitosan *in vitro* (Dvorackova 2011; Table 1).

**Table 1: Simulating GIT conditions**

Part of GIT	pH	Medium	Time
Stomach	3.0	Phosphate buffer	2 h
Small intestine	6.8	+ 4.2 g Na <sub>3</sub> PO <sub>4</sub> . 12 H <sub>2</sub> O	3 h
Terminal ileum	7.5	+ 6.0 g Na <sub>3</sub> PO <sub>4</sub> . 12 H <sub>2</sub> O	0.5 h
Colon	4.0/ 6.0	Acetate buffer/ phosphate buffer + $\beta$ -glucosidase	16.5 h

### Animal studies

Animal experiments (*Guide for the Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council and National Academy of Sciences, USA) with male Wistar rats (175-199 g; Société Janvier, Le Genest, France) in 5 groups (n=4) according to colitis and drug administration (healthy control group; colitis control group - non treated; rutin solution receiving group per rectum; rutin solution receiving group per os and rutin pellets receiving group per os) were provided after a week of acclimatization. Colitis was induced (Morris 1989) and developed for 2 days. Treated groups received rutin (10 mg/kg) once daily for 5 continuous days. Colitis controls received physiological saline orally instead. One day after the treatment, the animals were sacrificed and colons were resected. Colon/body weight ratio (Lamprecht 2005) and myeloperoxidase activity were determined (Krawisz 1984).

## RESULTS AND DISCUSSION

### Pellet evaluation *in vitro*

Pellets of very good characteristics: pellet mean diameter 0.77 mm, sphericity 0.83, hardness 9.35 N, friability 0.21 % and Hausner ratio 1.08, were obtained. Rutin content was 29 %, i.e. 97 % of its theoretical value. Coated pellets showed low rutin dissolution (14 %) under upper gastrointestinal tract conditions, while its release was fast (close to 90 %) under the conditions mimicking the colon; that is a good presumption of intended rutin release (Fig. 1).

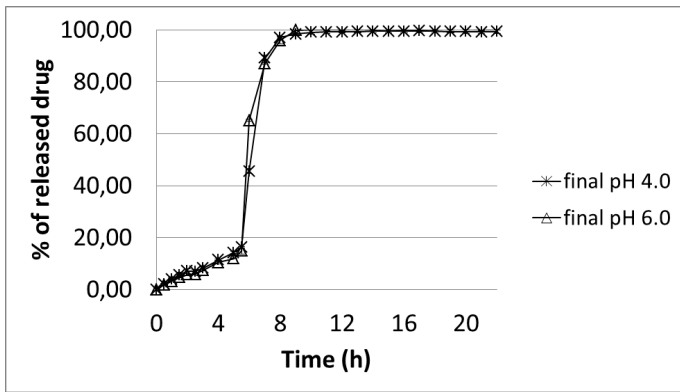


Figure 1: Rutin dissolution profiles

**In vivo studies**

In vivo experiment, rutin was able to promote colonic healing at the dose of 10 mg/kg, i.e. colon/body weight ratio decreased (Fig. 2) and myeloperoxidase activity was significantly suppressed (Fig. 3). The best colon/body weight ratio results (Fig. 2) were obtained in group receiving pellets (0.008) or rutin solution rectally (0.009). In healthy control group, colon/body weight ratio was also calculated and reached the value of 0.004. Concerning myeloperoxidase activity (Fig. 3), coated pellets showed excellent therapeutic efficiency and the value of 250 U/g tissue, i.e. myeloperoxidase activity reduction almost to the value of the control group.

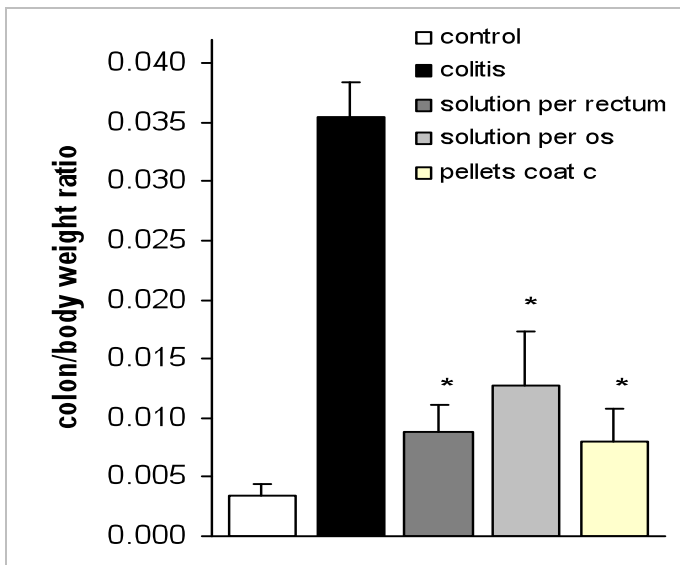


Figure 2: Colon/body weight ratio

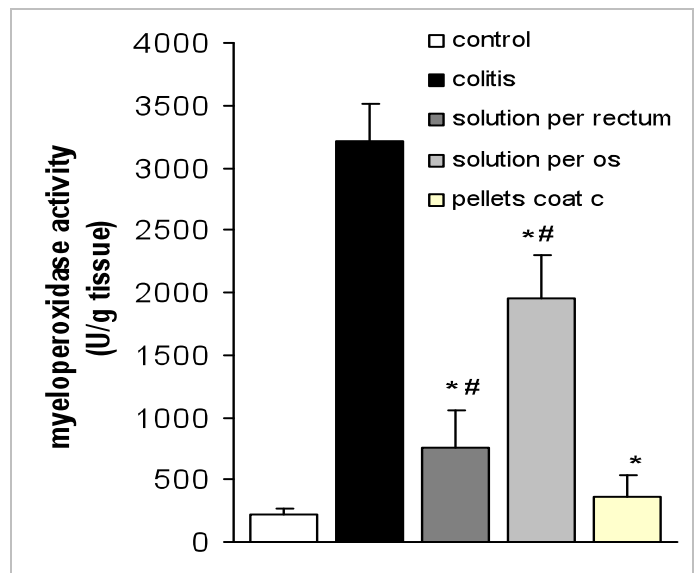


Figure 3: Myeloperoxidase activity

**CONCLUSIONS**

The combination of natural rutin, mucoadhesive chitosan degraded in the colon together with alginate coating, and pellets as multiparticulate dosage form having numerous advantages in IBD treatment could form a promising preparation free of side effects for lifelong therapy of this severe illness.

**REFERENCES**

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