

Hofmeister ions series protected protein during contact with organic interface

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INTRODUCTION

The association between chitosan and liposomes obtained by Reverse phase evaporation method (REVs-Chi) stabilized by polyvinyl alcohol (PVA) is proposed as a controlled delivery for Diphtheria toxoid (Dtxd). During this process, the protein suffers stresses with the organic solvent contact and the strong agitation.

Because preparation method characteristics a pre-formulation study on the Dtxd structure is shown.

The protein integrity after sonication with ethyl acetate in the presence of Hofmeister ions series salts was followed by HPLC^{2,3}, CD, ELISA and fluorescence³.

MATERIAL AND METHODS

Materials:

Dtxd (Instituto Butantan); all salts and organic solvents were of analytical grade.

Methods:

REVs-Chi preparation:

Briefly, the Dtxd in 0,5 % chitosan was added to soy phosphatidylcholine/cholesterol/ethyl acetate. After micelle formation, by sonication, the formed organogel was resuspended in PBS or 1% PVA¹.

Dtxd stability:

The Dtxd was diluted in KSCN; NaH₂PO₄; NaCl or MgCl₂ (0 to 150 mM salt) at fixed protein concentration (85,6 μM), and then added to a fixed volume of CH₃CO₂C₂H₅ (ethyl acetate). Immediately the emulsions were sonicated at 40 W during 4 minutes. The emulsions were centrifuged and the aqueous phases containing the soluble protein were analyzed by HPLC^{2,3}, ELISA, UV, fluorescence and CD spectroscopies.

RESULTS AND DISCUSSION

Analysis of Dtxd solubility and immunological identity

Dtxd remained 98% soluble in the presence of PO₄²⁻ and decreased (5-40 %) in the presence of Cl⁻, SCN⁻ and Mg²⁺ (Figure 1).

PO₄²⁻ retained the Dtxd conformation immunologically recognized (Figure 2).

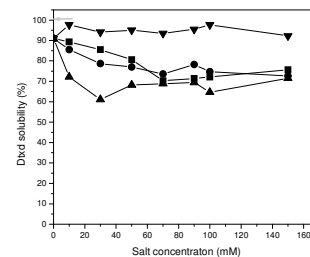


Figure 1. Effect of Hofmeister ion series salts in the Dtxd solubility. The protein's concentrations were calculated by absorbencies at 280 nm in the presence of (■) NaCl, (●) KSCN, (▲) MgCl₂ e (▼) NaH₂PO₄, after sonication with ethyl acetate (CH₃CO₂C₂H₅), by UV spectroscopy. Control: not sonicated Dtxd in water (→).

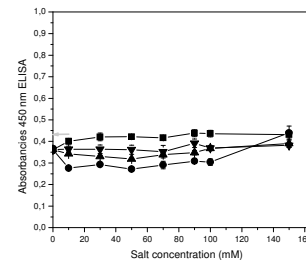


Figure 2. Effect of Hofmeister ion series salts in the immunological Dtxd identity after sonication. The Dtxd soluble fractions in the presence of (■) NaCl, (●) KSCN, (▲) MgCl₂ e (▼) NaH₂PO₄. Control: not sonicated Dtxd in water (→).

Analysis of concentration of molecular species after sonication by HPLC

It was not observed formation of insoluble aggregates in the presence of all studied salts (Figure 3).

NaH₂PO₄ is the salt that provided more protection of protein (Dtxd): this salt increased monomer concentration and decreased fragment A and B concentration compared to the other studied salts (Figure 3).

KSCN increased the FA and FB (Dtxd peptides) contents. It means: this salt induced protein damages (Figure 3).

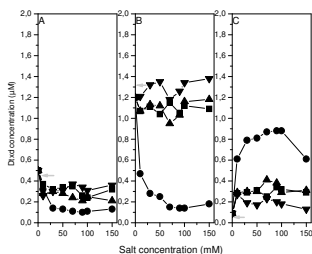


Figure 3. Effect of Hofmeister ion series salts in the Dtxd concentration of molecular species after sonication. The Dtxd samples were sonicated with Ethyl acetate ($\text{CH}_3\text{CO}_2\text{C}_2\text{H}_5$) in the presence of (-■-) NaCl, (-●-) KSCN, (-▲-) MgCl_2 e (-▼-) NaH_2PO_4 . The samples containing 2 μM Dtxd were analyzed by HPLC gel filtration (flow = 0,6 mL/minute in PBS pH 7,2, 20°C). A. Dimmer concentration, B. Monomer concentration, C. Fragment A and B concentration. Control: not sonicated Dtxd in water (→).

Analysis by Fluorescence and Circular Dichroism

W intrinsic fluorescence: The tertiary Dtxd structure remained stable (Figure 4).

The Dtxd β -sheet content remained constant in the presence of all the studied salts (Figure 5).

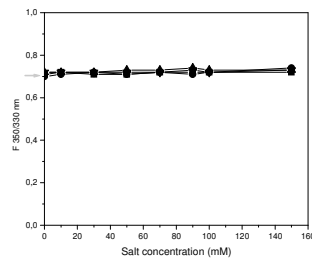


Figure 4. Relation between intrinsic fluorescence at 350 and 330 nm of Dtxd samples in the aqueous phase after sonication. The samples (-■-) NaCl, (-●-) KSCN, (-▲-) MgCl_2 e (-▼-) NaH_2PO_4 after sonication with ethyl acetate ($\text{CH}_3\text{CO}_2\text{C}_2\text{H}_5$) were analyzed by fluorescence spectroscopy. Control: not sonicated Dtxd in water (→).

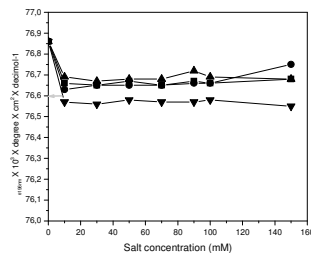


Figure 5. Effect of Hofmeister ion series salts in the Dtxd in the $[\theta]196$ nm. The Dtxd samples (7,36 μM) in the presence of (-■-) NaCl, (-●-) KSCN, (-▲-) MgCl_2 e (-▼-) NaH_2PO_4 , after sonication with ethyl acetate ($\text{CH}_3\text{CO}_2\text{C}_2\text{H}_5$), were analyzed by CD spectroscopy. Control: not sonicated Dtxd in water (→).

The Dtxd α -helix content remained constant in the presence of PO_4^{2-} (Figure 6).

The S-S dihedral angle remained stable in the presence of all studied salts (Figure 7).

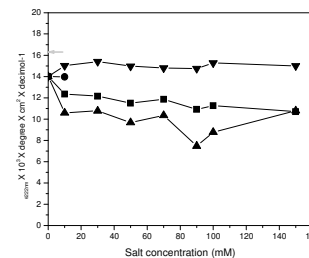


Figure 6. Effect of Hofmeister ion series salts in the Dtxd in the $[\theta]222$ nm. The Dtxd samples (7,36 μM) in the presence of (-■-) NaCl, (-●-) KSCN, (-▲-) MgCl_2 e (-▼-) NaH_2PO_4 , after sonication with ethyl acetate ($\text{CH}_3\text{CO}_2\text{C}_2\text{H}_5$), were analyzed by CD spectroscopy. Control: not sonicated Dtxd in water (→).

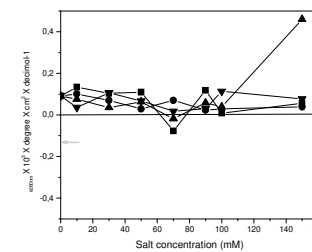


Figure 7. Effect of Hofmeister ion series salts in the Dtxd in the $[\theta]260$ nm. The Dtxd samples (7,36 μM) in the presence of (-■-) NaCl, (-●-) KSCN, (-▲-) MgCl_2 e (-▼-) NaH_2PO_4 , after sonication with ethyl acetate ($\text{CH}_3\text{CO}_2\text{C}_2\text{H}_5$), were analyzed by CD spectroscopy. Control: not sonicated Dtxd in water (→).

CONCLUSIONS

The Hofmeister salt series protected Dtxd during the REVS-Chi preparation. The Dtxd α -helical content was quite stable with minor exposition of hydrophobic residues (corroborated by measures on intrinsic tryptophan (Trp) fluorescence). REVS-Chi liberated soluble and immunological active Dtxd. The formulation can be used as vaccine particulate adjuvant.

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